

Short Communication

A rapid alternative technique for obtaining silver-positive patterns in chromosomes

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Abstract

Silver nitrate chromosome staining to evidence nucleolar organizer regions (NORs) is a widely adopted methodology. The aim of the present work was to improve this technique, reducing the preparation time without decreasing the quality of the results. Microwave irradiation proved to be quite efficient and reliable for this purpose, as it allowed to identify Ag-NORs equivalent to those obtained by the conventional procedure and also to reduce the concentration of the employed reagents, as well as the precipitation of debris on the preparation.

Key words: silver staining, Ag-NOR, microwaves, chromosome banding.

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Introduction

The chromosomal localization of 45S (18S+5.8S+28S) and 5S ribosomal genes can be accomplished by fluorescent *in situ* hybridization (FISH) with specific rDNA probes (Pinkel *et al.*, 1986). However, the most commonly adopted method for detecting nucleolar organizer regions -NORs - has been silver nitrate staining, because of its efficacy and easy performance (Goodpasture & Bloom, 1975; Howell & Black, 1980; Rufas *et al.*, 1982).

It is assumed that silver nitrate attaches to nucleolar proteins and not directly to rDNA (Miller *et al.*, 1976; Goessens, 1984; Hernández-Verdún, 1986), besides the necessity of transcriptional 45 rDNA activity during the preceding interphase (Howell, 1977; Hubbell, 1985; Jiménez *et al.*, 1988). Nevertheless, it is also hypothesized that the uncondensed state of the chromatin could be enough to determine chromosome impregnation by silver nitrate, thus discarding the requirement for transcriptional pre-activity (Clavaguera *et al.*, 1983; Medina *et al.*, 1983; Sánchez-Pina *et al.*, 1984). In addition, rDNA-free regions can also be evidenced by silver staining, probably due to the presence of proteins rich in acidic residues (Sumner, 1990; Sanchez *et al.*, 1995; Dobigny *et al.*, 2002).

The localization of NOR sites is an important tool in certain studies, such as those on evolution and cytotaxonomy (Galetti Jr., 1998), and those on gene expression and diagnosis of pathogenic tumors (Pich *et al.*, 2000). In the latter, the mean number of Ag-NORs is used as a patho-

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logical marker for some malignant tumors. As an alternative to the standard procedures, it was suggested to utilize microwave irradiation for nucleolus localization in histological preparations, in order to fasten and improve the diagnosis (Medina *et al.*, 1995; Li *et al.*, 1995).

The objective of the present work was to test the viability of microwave utilization to detect Ag-NORs in conventional chromosome preparations, obtained by airdrying from a cell suspension.

Material and Methods

Chromosome preparations of *Leporinus mormyrops* (Pisces: Anostomidae) were obtained by cell suspension technique (Bertollo *et al.*, 1978). Control slides were stained with silver nitrate, according to the conventional procedure described by Howell & Black (1980).

Onto the test slides, two drops of 1% aqueous gelatin solution with 0.25% formic acid and four drops of silver nitrate at 25% were placed. The slides were covered with coverslips and incubated for five seconds in the presence of high-potency microwaves. After incubation, the coverslips were removed and the slides were washed under tap water, stained with 5% Giemsa for 30 s, and air-dried.

The slides were analyzed under an optical microscope, and the selected metaphases were captured by using the Media Cybernetics image analysis system with the Image Pro Plus software.

Results and Discussion

The species *Leporinus mormyrops* was chosen because it bears a single pair of Ag-NORs. The results ob-

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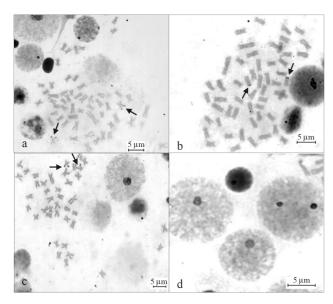


Figure 1 - Nucleolar organizer regions after silver nitrate staining using microwaves. (a-c) Metaphases. The arrows indicate the NORs sites. (d) Interphase nuclei.

tained are shown in Fig. 1 (a-c). It was observed that the quality of Ag-NORs revealed by the method described herein was similar to that obtained by the conventional method. Also, the nucleoli of interphase nuclei were quite evident (Fig. 1-d).

In the conventional method of Howell & Black (1980), the silver nitrate impregnation has to be monitored in a drying chamber, until the adequate staining is obtained. At a temperature of 60 $^{\circ}$ C, this time can vary from 3 to 5 min (in the drying chamber); higher temperatures usually lead to excessive drying, decreasing the preparation quality.

In the diagnosis of pathogenic tumors, the utilization of microwave irradiation shortened the time needed to evidence the nucleoli (Medina *et al.*, 1995; Li *et al.*, 1995) by up to one minute or close to it. Similarly, the exposure of the slides to microwaves drastically reduced the time needed for Ag-NOR detection, as they could be evidenced in approximately 5 s.

The utilization of lower concentrations of both gelatin and silver nitrate is required to avoid the preparations to dry up, since the mechanism of action of microwaves in conventional ovens is based on the agitation of water particles, producing heat. Additional Giemsa staining improves the contrast between chromosomes and Ag-NORs, therefore being helpful for metaphase analysis.

Besides saving time, as compared to the usual silver nitrate staining technique, and eliminating the constant monitoring until an adequate coloration is reached, this procedure also allows to prepare several slides at the same time, as the microwave irradiation is homogeneously distributed within the oven. Another advantage of this method is the reduction of silver debris precipitation, common in the conventional procedure and responsible for difficulties in the analysis of the preparations.

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