PREPARATION OF HYBRID POLYMERS AND USE AS CONTROLLED RELEASE DELIVERY VEHICLE FOR VOLATILE REPELLENT TO CONTROL *Aedes aegypti*

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The Zika virus is a flavivirus spread by the *Aedes aegypti* mosquito, which also transmits dengue and chikungunya. Current forms of control include elimination of mosquito breeding sites, use of insecticides, repellents and other means. The present study developed new adsorbent materials based on silica and biomaterials capable of releasing an *Aedes aegypti* mosquito repellent. The materials synthesized were characterized by Fourier-Transform Infrared, thermogravimetric analysis, Scanning Electron Microscopy and Brunauer, Emmett and Teller analysis and tested with citronella, an essential oil widely recognized as a natural insect repellent. The materials presented type IV isotherms, with type H1 hysteresis, characteristic of mesoporous materials, with pore diameters of between 50 Å and 80 Å. Materials based on silica-chitosan-glutaraldehyde dried in an oven (SCGO) and silica-chitosan-glutaraldehyde dried under a vacuum (SCGV) were found to produce more controlled release of citronella essential oil. The bioassay test was performed using a horizontal double-choice olfactometer. The adsorbent SCGO remained stable for the four hours of the bioassay, while the commercial repellent became ineffective after two hours of release. The production of a prototype in the form of bracelets, stickers releasing a volatile repellent is therefore proposed as a method for the control of the mosquito in urban areas.

Keywords: adsorbents; volatile repellents; biopolymers.

INTRODUCTION

The *Aedes* (*Stegomyia*) *aegypti* Linnaeus mosquito is a cosmopolitan species that is present mainly in tropical and subtropical regions, where climatic and social conditions favor its development. It is found in all the States that make up the Brazilian Federative Republic and in approximately 4,500 municipalities in Brazil. For reason of its extensive global distribution, it is involved in the transmission of various arbovirus diseases, such as Zika, dengue, yellow fever and chikungunya.¹

The Zika virus is a flavivirus (Flaviviridae family) that was first isolated from a febrile female Rhesus monkey in the Zika Forest, located near Entebbe in Uganda, from which the virus takes its name.² This virus is related to the yellow fever and dengue virus, which are also transmitted by the Aedes aegypti mosquito and cause hemorrhagic fever. The Zika virus causes fever, accompanied by other mild general symptoms, such as headache, rash, malaise, edema and sometimes severe joint pain. However, despite the apparent benign nature of this disease, more severe conditions, including central nervous system involvement (Guillain-Barré syndrome, transverse myelitis and meningitis) have recently been reported to be associated with Zika in French Polynesia and Brazil. This shows how little is known about this disease. The World Health Organization declared the Zika virus outbreak in the Americas a global emergency. This followed the rapid emergence of the virus in Brazil and the South American subcontinent and recognition of a link with the rise in the number of babies born with microcephaly and other neurological abnormalities.³

Dengue fever has been a public health problem in Brazil for many years. Since 2015, Zika and chikungunya have also emerged and have caused deaths and serious health problems among the Brazilian population. The main strategy adopted to combat these diseases is extensive use of insecticides and larvicides. These, however, have been shown to be ineffective and are also toxic. There is hope of development of a vaccine against Zika, but it may take years for such a vaccine to be available for use. In consequence of this, there is an ongoing search for new simple fast strategies for elimination of the Zika vector mosquito, since many deaths have been registered, and the quality of life of many impaired. This is especially the case among those afflicted with chikungunya, which can remain debilitating for months or even years after initial infection.

In view of the damage to human health caused by *Ae. aegypti*, numerous studies have been conducted to find alternative ways to control this vector. These include the use of plant repellents, to prevent mosquito bites and consequently the transmission of the arboviruses that cause disease.⁴⁻⁶

The degree of efficacy of repellents depends greatly on the composition of the product. Formulations based on creams, polymer blends or microcapsules for controlled release increase the duration of repellence activity.⁷ Polymers, such as activated carbon, Porapak Q, Tenax GR or Tenax TA, are used in analyses of the retention of volatile organic compounds.^{8,9} Chitosan is another polymer that is widely used in research, as it possesses excellent biodegradation properties and is a naturally occurring polysaccharide.¹⁰ This substance has produced excellent results against insects, with studies showing that the most active derivative of chitosan (N- (2-chloro-6-fluorobenzyl), when incorporated into an artificial diet, caused 100% mortality of larvae.¹¹ Nanoparticles of chitosan synthesized from shrimp and crab exoskeletons incorporated in a solution tested in a larvicidal bioassay against *Ae. aegypti* have produced mortality of 98% and 93% respectively at a concentration of 100 mg. L^{-1,12}

Chitosan has also been used with technology relating to the microencapsulation of the essential oils of plants, to reduce loss through evaporation. For reason of its high volatility, this technique is commonly used in the processing of repellents containing citronella essential oil (*Cympobogon* spp.) Organic-inorganic hybrids are materials formed by the combination of organic and inorganic components that produce materials with characteristics different from those of their components but that reflect the chemical nature of both, combining the thermal stability of inorganic materials with the chemical characteristics of organic compounds.¹³ The present study

thus aimed to develop *Aedes aegypti* control strategies that are less harmful to the environment and present greater stability.

EXPERIMENTAL

The present study synthesized the adsorbents, investigated their adsorptive properties, and measured citronella essential oil release. All reagents used were of analytical grade and deionized water (18 M Ω cm) was obtained using a Millipore Inc Milli-Q purification system.

Synthesis of silica-chitosan-glutaraldehyde

For the preparation of the SCG material, 2.0 grams of chitosan were solubilized in 500 ml of 2% (v / v) acetic acid. Ten grams of silica gel were then added to the chitosan solution, followed by 100 ml of glutaraldehyde 20%. After filtration, the solid material was washed with deionized water to remove excess glutaraldehyde, leaving no trace of this compound. The material was then divided into two fractions and dried using two different methods. The first fraction was distributed in a Petri dish, which was subsequently placed in a desiccator with a suction pump for 24 hours. This was called silica-chitosan-glutaraldehyde dried under vacuum (SCGV). The second fraction was heated in an oven at 50 °C for 24 hours. This fraction was called silica-chitosan-glutaraldehyde dried in an oven (SCGO).

Characterization

The synthesized materials (SCGV and SCGO), chitosan, silica gel and glutaraldehyde were characterized by Fourier Transform Infrared Spectroscopy (FTIR) according to the methodology adapted by Silva *et al.*¹⁴ in order to identify organic functional groups that may act as adsorption sites. The areas of the materials were determined using the method developed by Brunauer, Emmett and Teller (BET), whereby the adsorption and desorption isotherms are calculated at the temperature of liquid nitrogen (-196 °C). The FTIR spectra were obtained by attenuated total reflection (ATR) carried out at the Center for Analysis of the UFAL's Institute of Chemistry and Biotechnology.

Thermogravimetric (TG) analyses were also performed to evaluate the thermal behavior of the adsorbent materials.¹⁵ These analyses were performed in a temperature range of 25-800 °C with a heating rate of 10 °C min⁻¹ and a dynamic nitrogen atmosphere (20 mL min⁻¹) at the Catalysis and Fuels Laboratory of the UFAL's Department of Chemical Engineering. Finally, the physical structure of the material was determined using a Scanning Electron Microscope (SEM) in order to observe the morphology and any chemical or physical modification of the adsorbents.

Tests of release of citronella essential oil

The citronella essential oil was first placed beneath the materials using an automatic pipette and analytical balance, to obtain a concentration of 5% (m/m). The release studies were performed using thermogravimetric (TG) analyses with a 40 °C isotherm and a dynamic atmosphere of 20 mL / min of nitrogen. The release test was also performed using an analytical balance over a period of eight days.

Y-tube olfactometer bioassays

The durability of the behavior of mosquitoes in response to repellent treatments was examined in approximately 15-day-old laboratory-bred *Aedes aegypti*, kept at a uniform temperature of 27 ± 2 °C. This analysis was carried out using an olfactometer in the form of a Y-glass tube (trunk, 340 mm, arms, 290 mm, 45° angle, internal diameter, 40 mm) connected by the arms to silicone hoses connected to a fluxmeter, as shown in Figure 1. The air flow inside the olfactometer, purified by the active carbon filter, was 1200 mL min⁻¹. The experiment was carried out in a room with ambient temperature of 27 ± 2 °C and relative air humidity of around 80%. It should be noted that the whole system was cleaned with a neutral detergent and distilled water, followed by an ethanol bath, thus ensuring the removal of any impurity or odor that could possibly interfere with the experiment.

The adsorbent SCGO material containing 5% (m/m) of the citronella essential oil was placed in one of the Y tube connections, and the control, without citronella, as well as the test with the citronella-based commercial repellent at a concentration of 5% (m/m) were placed in the other connection. After installation of the entire olfactometry unit, ten mosquitoes were inserted into the trunk of the olfactometer and these individuals were observed for four hours, in accordance with the protocol developed by Oyarzún *et al.*¹⁶ This process was performed in quadruplicate.

Only situations where the mosquito lasted for more than two hours were counted as a positive response. The results were fed into the following equation developed by the WHO:¹⁷



Figure 1. Representation of the Bioassay experiment



Figure 2. Nitrogen adsorption-desorption isotherms of the synthesized materials (a) SCGV and (b) SCGO

$$IA(\%) = 100 \times \left[\frac{Ca - Ta}{Ca}\right]$$
(1)

where IA is the percentage of landing inhibition, Ca is the number of mosquitoes that land in the control space and Ta is the number of mosquitoes that land in the treatment space. The results obtained by the equation were submitted to analysis of variance (ANOVA), followed by the Tukey test with a 5% level of significance, using the GraphPad Prism 6.0 Program.

 Table 1. BET surface areas and pore volumes of the SCGV and SCGO materials

Sample	S (m ² g ⁻¹)			V(cm ³ g ⁻¹)		
	S _{BET}	\mathbf{S}_{Ex}	S _{Micro}	V _{Micro}	$V_{\scriptscriptstyle Total}$	
SCGO	125	183	153	0.0329	0.2305	
SCGV	107	167	133	0.0344	0.1927	

 S_{BET} : Area by the BET method; S_{Es} : External area, S_{Micro} : Micropore area; V_{Micro} : Micropore volume; V_{Total} : Total volume of pores.

RESULTS AND DISCUSSION

The Brunauer, Emmett and Teller Method

The surface area, pore volume and distribution of pore sizes for the SCGV and SCGO materials were measured using the Brunauer, Emmett and Teller method (BET) (Figure 2).

The data on the BET-surface areas, pore volumes and pore sizes of the SCGV and SCGO are presented in Table 1. It can be seen that a there was a small variation in surface area according to the drying method used (oven drying or vacuum). There were almost no changes in pore volume. The adsorbents presented pore diameters of between 50 Å and 80 Å. According to Sing and colleagues adsorbents with pore diameter of 20 and 500 Å can be classified as mesoporous materials.¹⁸

The isotherms thus presented characteristics of type IV, with hysteresis of type H1, with the lower branch representing the quantity of gas adsorbed by increasing relative pressure, while the upper branch represents the amount of gas desorbed by the inverse process. Hysteresis is associated with the differences between the adsorption and desorption processes. Hysteresis type H1 is characteristic of adsorbents with a narrow and relatively uniform pore distribution.¹⁹

Spectroscopy in the Fourier-transform infrared region

Table 2 shows the main vibrational bands of the FTIR spectra of the compounds: (a) Silica, Chitosan, Glutaraldehyde and SCGO; and (b) citronella essential oil (3,7-dimetyl 2,6-octadienal). The silica gel spectrum showed bands at 969 and 1.096 cm⁻¹, related to Si-O stretch.²⁰ The chitosan spectrum presented bands at 1.560 cm⁻¹, attributed to the deformation of the NH2 group of the chitosan, and bands at 2.870 cm⁻¹ attributed to the CH stretch of the chitosan. The bands at 1.030 and 1.150 cm⁻¹ are related to the C-O stretch present in the structure. The SCGO spectrum shows the presence of the silica and chitosan bands with small displacements, indicating an interaction between the functional groups present in these compounds and the formation of a new material. The bands at 1.325 and 1.375cm⁻¹ relate

to the stretching of the amide type, which is formed in the reactions of the amine groups of the chitosan with the glutaraldehyde.²¹

Table 2. Main vibrational bands of the silica, chitosan, glutaraldehyde and

 SCGO materials

Material v(cm ⁻¹) Attribution		Attribution of the Main Vibrational Bands		
	969	Si-O Stretching		
Silica	1069	Si-O Stretching		
	2870	C-H Stretching of the Chitosan		
	1560	Deformation of the NH ₂ group		
Chitosan	1030	C-O Stretching		
	1150	C-O Stretching		
Glutaraldehyde	2944	C-H Propyl Stretching		
	1069	Si-O Stretching		
	1375	Amide Stretching (reaction with glutaraldehyde)		
SCGO	1030	C-O Stretching		
	1150	C-O Stretching		
	2870	C-H Stretching of Chitosan		
	2944	C-H Propyl Stretching		

The citronella essential oil spectrum, Figure 3, thus shows bands at 2.965 and 2.917 cm⁻¹, indicating the presence of the CH₂ group and CH₃ aliphatic, as well as the band at 1.455 cm-1 characteristic of the angular deformation (CH₂)n, when n < 3. It also presented the band at 1.377 cm⁻¹ characteristic of the germinal dimethyl angular deformation (CH₃) and the band at 1.641 cm⁻¹ characteristic of alkene stretching (-C=C-). As an aldehyde, citronella essential oil also shows the bands characteristic of this functional group, such as CH stretching of the aldehyde at 2.715 cm⁻¹ and the band characteristic of the carbonyl group (C=O) of aldehyde at 1.720 cm⁻¹.²²

We also found, using the FTIR technique, that there was an

Scanning electron microscopy

Scanning electron microscopy (SEM) was used for morphological characterization of chitosan, SCGO and SCGV (Figure 5). These images show a clear difference between the chitosan and the prepared materials, with the chitosan exhibiting a rough surface and more porous appearance compared to the other materials. Generally, speaking particles of non-uniform size are found in hybrid materials, as is the case with silica gel. The incorporation of chitosan reduces the amount of empty space between particles. On the other hand, the surface of the SCGO and SCGV materials were found to be similar, as is to be expected when the materials are the same, differing only with respect to the drying method used.

Thermogravimetric analysis

The thermic behavior of the chitosan, SCGO and SCGV materials was evaluated by thermogravimetric analysis. It has been reported in the literature that the hybrid compounds formed by chitosan and silica present a three-stage pattern of thermal decomposition. The thermogravimetric (TG) and first derivative (DTG) curves for the synthesized materials were obtained and the data on loss of mass are presented in Table 3. TGA results for the chitosan and chitosan with citronella are shown in Supplementary Material. For chitosan alone, there is an initial mass loss of 5% at 100 °C related to water loss. In the presence of the oil produced 36% weight loss in the same temperature range. It is interesting to note that comparing both materials can be observed a weight loss 31% related to citronella oil adsorbed or absorbed in the chitosan (310 mg g⁻¹).

Evaluation of the TGA and DTG curves of the synthesized materials revealed no differences, suggesting that the materials have similarities with respect to their chemical composition. The

Citronella

1000

500

 Wave numbers (cm⁻¹)
 Wave numbers (cm⁻¹)

 Figure 3. Spectroscopy in the infrared region of the compounds: (a) Glutaraldehyde, Silica, Chitosan and the SCGO material, (b) Citronella essential oil

120 b) 140 a) SCGV SCGO 130 SCGV+Citronella SCGO+Citronella 120 Transmitance(%) 110 Transmitance(%) 90 100 271 90 2715 80 291 60 70 296 60 50 30 3500 3000 2500 2000 1500 1000 500 3500 3000 2500 2000 1500 1000 500 Wave numbers (cm⁻¹) Wave numbers (cm⁻¹)

Figure 4. Spectroscopy in the infrared region for the compounds: (a) SCGV and (b) SCGO, in the presence of citronella at 5%, after 8 days of release

Figure 5. SEM of (a)Chitosan 1000x; (b) SCGO 1000x; (c) SCGV 999x





	1 st mass loss		2 nd mass loss		3 rd mass loss		
Material	Mass loss (%)	Temperature range (°C)	Mass loss (%)	Temperature range (°C)	Mass loss (%)	Temperature range (°C)	TOTAL
SCGV	40	40-200	15	200-260	10	300-500	65%
SCGO	35	40-200	15	200-260	10	300-500	60%

Table 3. Data on the mass loss of materials

first stage of mass loss occurred between 40 and 200 °C. This event was responsible for the mass loss of 30% to 45% of the material, as confirmed by the DTG curve, in which the temperature at which this phenomenon occurs is 135 °C. This can be attributed to the release of residues of acetic acid, glutaraldehyde and citronella physically adsorbed on the surface of the materials.

It is also possible that the silica itself contributes to the initial mass loss peaks of these materials, as a result of residual silane groups present on the surface of the silica gel that have not undergone a reaction, enabling these groups to cause each other to condense, releasing water as volatile product.²³

The SCGV material presented a larger quantity of volatiles, citronella and adsorbed water, and a greater initial loss of mass was consequently observed. This is caused by the oxygenated and nitrogenous groups present in chitosan, which interact with water through hydrogen bonds. The second mass-loss event corresponds to the decomposition of the chitosan present in large quantities in this material due to the glutaraldehyde cross-linking. In the third loss of mass, the event is attributed to some change in the silica due to the high temperature, presenting a loss of 10% up to a temperature of 500 °C. It is also possible that a decomposition phase occurs, through the dehydration of saccharide rings, depolymerization and decomposition of the organic polymer units.²⁴

Evaluation of the DTA graphs for the SCGO and SCGV materials reveals two distinct events related to mass loss: one with an endothermic profile with a maximum of 135 °C, related to the evaporation of the volatile compounds and another with an exothermic profile, with a maximum of 260 °C for the degradation of the cross-linked chitosan (Supplementary Material).

Study of the citronella essential oil release

The volatilization characteristics of the essential oil adsorbed by different materials were studied using an isothermal system. The mass loss and mass loss rate of citronella for Chitosan and the hybrid material were determined using thermogravimetric analysis similar to that used for analysis of coal and these figures were used to determine release constants.

Initially, the sample was prepared by adding 8 mg of citronella to 20 mg of the adsorbent material. Citronella thus accounted for 28.6% of the mass of the sample. The studies of citronella essential oil release were performed using isothermal gravimetric analysis at a temperature of approximately 40 °C (Figure 6a). For chitosan, citronella desorption was greater than for the hybrid material. For the hybrid materials (SCGO and SCGV), the process was found to be linear from 2,000 seconds onwards. The DTA graph showed an event with endothermic characteristics up to 2,000 seconds, indicating that the volatile repellent is absorbed by the material. However, as our interest is in adsorbed content, a curve fitting was obtained for 2,000 seconds onwards to ensure that no event influenced the linearity of the release curves.

Considering the curve only after 2,000 seconds, where the process of citronella desorption occurs in a more linear fashion, characteristic of zero-order reactions, the constant was calculated for all materials synthesized using the graph method (Figures 6b and 6c), where k corresponds to the slope of the curve.

$$[A] = -kt + [A]_0$$
(2)

The release rate at 40 $^{\circ}$ C for each material was calculated based on the mass of citronella adsorbed and whether release occurred between 2000 and 7000 seconds (0 to 3000 seconds for chitosan). For chitosan, citronella desorption was greater than for the hybrid material. For hybrid materials the release rate was similar (Table 4).

Analysis of the mass loss release curve in an analytical balance for 8 days revealed similar behavior. The curve became more linear from the 2^{nd} day onwards, a find which is related to the quantity of mass adsorbed by the material. Mass released before the 2^{nd} day represents non-absorbed compound. This is shown by the abrupt decrease in the curve at the beginning of the experiment. As it exhibited a similar release behavior and was obtained using a simpler drying process, only adsorbent material SCGO was used in the bioassay studies

Bioassay

At the beginning of the experiment, it was common for the mosquitoes to appear agitated. This was probably because of the release of the citronella (C) absorbed in the material, which tends to be released quickly at the beginning of the process, as observed in the TG analysis.

The results obtained in the present study demonstrate the efficiency of the SCGO material in the presence of citronella in inhibiting Ae. aegypti for several hours. Figure 7 shows that there were no significant differences between the treatments used for the period of one hour, indicating that all treatments are initially efficient repellents. However, after two hours of bioassay, there was a significant difference between SCGO+C and the commercial repellent RC (P<0.01), as well as between SCGO+C and SCGO (P<0.05), but no difference between SCGO and RC. Behavior after two hours was found to be similar to that after two hours. It can thus be concluded that the commercial repellent and the product without citronella do not provide any increase in inhibition compared to the product containing citronella. After four hours of treatment, a significant difference was observed between the SCGO+C sample and the RC (P<0.01), as well as between SCGO and RC, but there was no significant difference between SCGO+C and SCGO. This suggests that the commercial repellent is not efficient.

The data obtained showed that, over the full four hours of the experiment, the SCGO+C treatment had a better rate of inhibition of the mosquito, with 100% inhibition in the first hour and 97.25% in the last. The SCGO treatment had the highest inhibition rate in the first hour of the experiment (72.91%), showing it to be the second best alternative for inhibition of *Ae. aegypti*, In the last hour there was a slight reduction in inhibition to 72.13% for this treatment. Although SCGO without citronella presented the second highest rates (in percentage) of inhibition to the mosquito, this treatment only differed from the commercial repellent in the last hour of the experiment,



Figure 6. a) Thermogravimetric release curves for the chitosan, SCGO, SCGV samples, with citronella; curves for the volatile repellent (2000-7000 seconds) with their respective linear regressions b) SCGO and C) SCGV

Material	Release rate (mg h ⁻¹) at 40 °C	Constant (%s ⁻¹)
Chitosan	0.94	-
SCGO	0.49	3.85×10^{-6}
SCGV	0.49	5.82×10^{-6}

in which it demonstrated greater efficacy. The commercial insect repellent was generally less effective than the other treatments, with inhibition rates of 63.09%, 66.33%, 46.66% and 36.66% for 1, 2, 3 and 4 hours duration, respectively, indicating a decline in efficiency over time. The results obtained for the SCGO material developed in the present study concur with the findings of Harismah *et al.* in a study involving a commercially available repellent paint with the addition of citronella oil and clover leaf, in which they found that, after a period of five hours, there was a tendency for efficiency to decrease.²⁵

According to Rodriguez *et al.*, the protection provided by a commercial repellent (RC) based on citronella oil (10%) has an estimated duration of two hours.²⁶ The same authors found that the repellents tested provided maximum protection for two hours. This was then followed by a gradual decline in efficacy up to the conclusion of the four-hour experiment. The findings of this study, however, show that the SCGO+C material is a more effective repellent even with a lower proportion of citronella (5%). Mendki *et al.* in bioassays to test the activity of formulated repellents containing 20% of a commercial synthetic repellent DEPA (N, N-diethyl phenyl acetamide) + one plant essential oil against *Aedes aegypti*, one of these being citrine, found that the average protection time for this formulation was 2.5 hours.²⁷

In a study of the effect of essential oils on the behavior of the Aedes aegypti mosquito involving an olfactometer bioassay, Uniyal et *al.* found that the essential oils of some plants such as litsea, rosewood and geranium were effective repellents of *Ae. aegypti*, but that these oils were not as effective as the synthetic insect repellents DEET (N,N Diethyl-m-toluamide) and DEPA.²⁸ These results contradict the results presented here, in which the commercial repellent was found to be less effective than the SCGO+C treatment.

CONCLUSIONS

The results show that the proposal for synthesis, characterization and study of the release of citronella essential oil by means of adsorptive processes using materials based on chitosan and silica gel was an efficient method. Through the characterization of techniques, it was possible to provide evidence of the formation of the desired materials, especially those relating to the incorporation of chitosan in silica through the glutaraldehyde crosslinking agent. The FTIR spectra and the thermogravimetric curves indicate the incorporation of clusters in the chitosan by way of glutaraldehyde crosslinking, and this produced the best adsorption capacity and release rate, in addition to a satisfactory release constant. In all materials, citronella was present even after eight days of release. SEM results indicate possible sites for adsorption.

The bioassays presented satisfactory results for controlled release of citronella essential oil by means of the adsorbent material SCGO, in which it was stable even after four hours of release.

It will thus be possible to develop a prototype in the form of bracelets or stickers based on the materials developed here, with a view to releasing these volatile compounds in a controlled manner and thereby helping to control the *Aedes aegypti* mosquito.

SUPPLEMENTARY MATERIAL

TGA/DTG and DTA curves for Chitosan, Chitosan + Citronella,



Figure 7. Bioassay using "Y" tube olfactometer to evaluate the duration of inhibition (%) of samples containing silica, chitosan and glutaraldehyde + citronella (SCGO+C) and SCGO against the Aedes aegypti mosquito compared to synthetic repellent (RC). P values: $P < 0.05^*$; $P < 0.01^*$; not significant (ns)

SCGV and SCGO are freely available at http://quimicanova.sbq.org. br, in PDF format.

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