

**SCHISTOSOMA MANSONI: IMMUNODEPRESSION OF
HEPATIC SCHISTOSOME GRANULOMA
FORMATION IN MICE INFECTED
BY TRYPANOSOMA CRUZI ***

O. Genaro¹, Z. Brener² and P.M.Z. Coelho³

Infection by Trypanosoma cruzi in mice depresses hepatic granuloma formation around Schistosoma mansoni eggs. This immunodepressive effect occurred in mice with Chagas' disease at the acute and/or chronic phases, granulomas being significantly smaller than those in controls. Data suggest that Chagas' disease depresses the delayed hypersensitivity immune response directly.

Key words: *Schistosoma mansoni*. Immunodepression. Granuloma. *Trypanosoma cruzi*.

Immunodepression during infection by *Trypanosoma cruzi* was first described by Clinton et al.⁵, who noted a reduction of early response of antibodies to donkey red blood cells in infected mice. Later, depression of immune response to different antigens in mice infected by *T. cruzi*^{17,18} and in men²² was demonstrated. The mechanisms of this immune depression in Chagas' disease have been studied by a number of authors^{6, 8, 21}. Macknika and Choromanski¹⁴ showed that *T. cruzi* induces nonespecific immunodepression in mice reducing humoral and cellular response to antigens of *Hymenolepis diminuta*. A marked depression of humoral and cellular response coincidentally with the peak of parasitemia occurs during the acute phase of infection by *T. cruzi*⁷.

In the present study we studied the effect of infection by *T. cruzi* (acute and chronic phases) on granuloma formation in mice with concomitant infection by *Schistosoma mansoni*.

MATERIAL AND METHODS

Swiss albino mice weighing 18-20 g were used. *S. mansoni* cercariae (LE strain, coming from Belo Horizonte, Brazil) were obtained from *Biomphalaria glabrata*^{13,16}. Mice were inoculated with about 40 cercariae subcutaneously. Three strains of *T. cruzi* were used: Y strain²⁰, CL strain⁴, and Colombian

strain¹². Induction of the chronic phase in CL and Colombian strains was performed according to Brener and Chiari³. An inoculum of about 1×10^4 blood trypomastigotes, given by the intraperitoneal route, was the standard challenge. Using these methods, granuloma formation was studied in two groups of mice: a) infected by both *S. mansoni* and *T. cruzi*; b) infected by *S. mansoni* only. Control groups of uninfected animals and those only infected with *T. cruzi* were maintained concurrently.

In experiment I, designed to study the effect of acute phase of Chagas' disease on schistosome granuloma formation, 20 mice were inoculated with *S. mansoni*. On the 43rd day after infection, 10 out of those 20 mice, plus 10 normal ones, were inoculated with *T. cruzi* (Y strain). Since this strain is highly virulent, with a high mortality rate after the peak of parasitemia, this experiment was of short duration.

In experiments II and III, mice chronically infected by *T. cruzi* (CL and Colombian strains), and their respective controls, were distributed in 4 different groups. At 98 and 185-day-intervals after infection by *T. cruzi* animals from groups A and C were infected by *S. mansoni*.

At the end of each experiment, surviving mice were killed and necropsy performed. Snips from the liver of infected mice were fixed in 10% formol-saline, embedded in paraffin, sliced into sections of 5 μ m thickness (with a 500 μ m interval), and stained with Hematoxylin-Eosin. The size of each granuloma was determined by the measure of two diameters intersecting at right angle, by means of an ocular micrometer.

RESULTS

Table I shows the mean diameter of schistosome granulomas in the liver of mice from group A (*S. mansoni* + *T. cruzi*) and group C (*S. mansoni* only) in

* This study was supported by CNPq and FINEP, Brazil.

1. Departamento de Patologia Tropical/INPA, Manaus

2. Centro de Pesquisas René Rachou, FIOCRUZ, Belo Horizonte

3. Departamento de Parasitologia, ICB/UFMG, Belo Horizonte

Address for reprints: Prof.º Paulo Marcos Z. Coelho

Departamento de Parasitologia, ICB/UFMG, Caixa Postal 2486, 30000 Belo Horizonte, MG.

Recebido para publicação em 26/6/85

Table 1 - Diameter of hepatic granulomas in mice with concomitant infection by *Trypanosoma cruzi* and *Schistosoma mansoni*

Phase of infection	Interval between infections (days)	Duration of infection by <i>S. mansoni</i> (days)	Duration of infection by <i>T. cruzi</i> (days)	Mean diameter of granulomas (μm)*		% reduction of granulomas in size	Student's <i>t</i> test	
				Group A T. cruzi+S. mansoni	Group B S. mansoni only			
I acute	Y	43	54	11	164.5 \pm 5.3	324.5 \pm 4.5	49.3	P < 0.001
II chronic	CL	68	62	130	195.7 \pm 5.9	368.8 \pm 11.1	46.9	P < 0.001
III chronic	Colombian	185	135	320	192.5 \pm 15.6	305.9 \pm 49.0	37.0	P < 0.01

* Mean diameter of granulomas of five mice (measured 40 granulomas per animal)

the three experiments. As can be seen, the diameters of granulomas in *T. cruzi* infected animals are significantly smaller than those in controls, with a reduction rate varying from 37 to 49.3% from the normal size of granulomas in control group.

DISCUSSION

Granulomatous reactivity around *S. mansoni* eggs has been demonstrated to be a form of delayed cell-mediated hypersensitivity²⁴. These authors showed that this kind of hypersensitivity could be transferable to non-infected mice with lymphnode cells or the spleen of immune animals, but not with serum. This hypothesis was reinforced by suppression of schistosome granuloma formation in mice, using effective techniques against delayed cell-mediated reaction, such as immunodepressive drugs¹¹, neonatal thymectomy⁹, heterologous antilymphocytic serum¹⁰ moderate or severe disease like Hodgkins²³, and antimacrophage serum². Abdel-Wahab et al.¹ first demonstrated that parasitic infections such as *Plasmodium yoelii* had direct influence on granulomatous reactivity around *S. mansoni* eggs reducing the size of granulomas. Later, Mahmoud et al.¹⁵ verified that infection by *Toxoplasma gondii* also induced a marked reduction of schistosome granuloma in size.

In the present investigation, infection by *T. cruzi* produced an intense effect on schistosome granuloma size. Depression of the delayed hypersensitivity immune response was showed by reduction of hepatic granulomas in size occurring in mice infected by *T. cruzi* (Y strain) at the acute phase. This finding agrees with those of Rowland & Kuhn¹⁹ and Costa⁷, who verified depression of cellular response by *T. cruzi* at the acute phase of infection. Further, it shows that such immunodepression also appears in Chagas' disease during the chronic phase, as observed with CL and Colombian strains of *T. cruzi*.

RESUMO

A infecção de camundongos pelo *Trypanosoma cruzi* inibe a formação do granuloma hepático esquistossomótico. Este efeito imunodepressor ocorreu em camundongos com a fase aguda ou crônica da doença de Chagas, sendo os granulomas significativamente menores que nos animais controles. Os dados sugerem que a doença de Chagas deprime diretamente a imunidade celular.

Palavras chaves: *Schistosoma mansoni*. Imunodepressão. Granuloma. *Trypanosoma cruzi*.

REFERENCES

1. Abdel-Wahab MF, Powers KG, Mahmoud SS, Good WC. Suppression of schistosome granuloma formation by malaria in mice. *American Journal of Tropical Medicine and Hygiene* 23:915-918, 1974.
2. Boros DL, Warren KS. Effect of anti-macrophage serum on hypersensitivity (*Schistosoma mansoni* eggs) and foreign body (Divinylbenzene copolimer bead) granuloma. *Journal of Immunology* 107:534-539, 1971.
3. Brener Z, Chiari E. Observações sobre a fase crônica da doença de Chagas experimental no camundongo. *Revista do Instituto de Medicina Tropical de São Paulo* 5: 128-132, 1963.
4. Brener Z, Chiari E. Variações morfológicas observadas em diferentes amostras de *Trypanosoma cruzi*. *Revista do Instituto de Medicina Tropical de São Paulo* 5: 220-224, 1963.
5. Clinton BA, Ortiz-Ortiz L, Garcia W, Martinez T, Capin R. *Trypanosoma cruzi*: Early immune responses in infected mice. *Experimental Parasitology* 37: 417-425, 1975.
6. Corsini AC, Costa MG. Immunosuppression in mice infected with *Trypanosoma cruzi* (Chagas, 1909). I. Evidence of polyclonal B cell activation in experimental infections mimiced by an extract prepared from circulating trypomastigotes. *Revista do Instituto de Medicina Tropical de São Paulo* 23:114-121, 1981.

7. Costa MC. Imunossupressão na infecção pelo *Trypanosoma cruzi* (Chagas, 1909): I. Efeitos dos extratos de epimastigotas e tripomastigotas sobre a resposta imune em camundongos. Tese de Doutorado. Unicamp. Campinas, 1982.
8. Cunningham DS, Benavides GR, Kuhn RE. Suppression of mitogen-induced blastogenesis by the *Trypanosoma cruzi*-induced suppressor substance. The Journal of Parasitology 66: 722-729, 1980.
9. Domingo EO, Warren KS. The inhibition of granuloma formation around *Schistosoma mansoni* eggs. II. Thymectomy. The American Journal of Pathology 51: 757-767, 1967.
10. Domingo EO, Warren KS. The inhibition of granuloma formation around *Schistosoma mansoni* eggs. III. Heterologous antilymphocyte serum. The American Journal of Pathology 52: 613-631, 1968.
11. Domingo EO, Cowan RBT, Warren KS. The inhibition of granuloma formation around *Schistosoma mansoni* eggs. I. Immunosuppressive drugs. The American Journal of Tropical Medicine and Hygiene 16: 284-292, 1967.
12. Federici EE, Albermann WB, Neva FA. Chronic and progressive myocarditis and myositis in C₃H mice infected with *Trypanosoma cruzi*. The American Journal of Tropical Medicine and Hygiene 13: 273-280, 1964.
13. Freitas JR. Ritmo de crescimento de *Biomphalaria glabrata* (Say, 1818). Padronização da técnica de criação. Tese de doutorado. Universidade Federal de Minas Gerais, Belo Horizonte, 1973.
14. Machnicka B, Choromawski L. The influence of immunosuppression generated by *Trypanosoma cruzi* on the development of *Hymenolepis diminuta* in CFW mice. Bulletin of l'Academie Polonaise de Science. 17: 739-748, 1979.
15. Mahmoud AAF, Strickland GT, Warren KS. Toxoplasmosis and the host-parasite relationship in murine schistosomiasis mansoni. Journal of Infectious Diseases 135:408-413, 1977.
16. Pellegrino J, Katz N. Experimental chemotherapy of schistosomiasis mansoni. In: Dawes B (ed) Advances in Parasitology, Academic Press, London, p. 233-290, 1968.
17. Ramos C, Lamoyi E, Feoli M, Rodrigues M, Perez M, Ortiz-Ortiz L. *Trypanosoma cruzi*: immunosuppressed response to different antigens in the infected mouse. Experimental Parasitology 45: 19-199, 1978.
18. Reed SE, Larson CL, Speer CA. Suppression of cell mediated immunity in experimental Chagas' disease. Zeitschrift fur Parasitenkunde 52: 11-17, 1977.
19. Rowland EC, Kuhn RE. Suppression of cellular responses in mice during *Trypanosoma cruzi* infection. Infection and Immunity 20: 393-397, 1978.
20. Silva LHP, Nussenzweig V. Sobre uma cepa de *Trypanosoma cruzi* altamente virulenta para o camundongo branco. Folia Clinica et Biologica 20: 191-207, 1953.
21. Teixeira ARL. Competência imunológica do paciente chagásico. Imunodepressão na forma aguda inaparente. Autoimunidade no hospedeiro imunizado. Tese de Doutorado. Universidade Federal de Minas Gerais. Belo Horizonte, 1981.
22. Teixeira ARL, Teixeira G, Macedo J, Prata A. Acquired cell-mediated immunodepression in acute Chagas' disease. Journal of Clinical Investigation 62: 1132-1141, 1978.
23. Warren KS. Inhibition of granuloma formation around *Schistosoma mansoni* eggs. The American Journal of Pathology 56:293-303, 1969.
24. Warren KS, Domingo EO, Cowan RBT. Granuloma formation around schistosome eggs as a manifestation of delayed hypersensitivity. The American Journal of Pathology 51: 735-756, 1967.